







1. 2018 short summary

Experiments/Collaborations
Trainings
Facilities

2. News for 2019 and development



Main Experiments/Collaborations using *Xenopus* oocyte system

In 2018: More than 30 xenopus surgeries for Functional studies

within BPMP (7 teams) and also with external collaborations







cRNA injection of protein of interest







Tracer uptake/efflux

Swelling

Electrophysiology

Hormones, metals, NO₃-, citrate ...

Aquaporins

Ion channels

A lot of studies/projects (Jeremy, Colette, ...): 2 examples

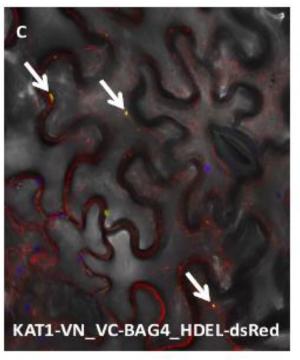


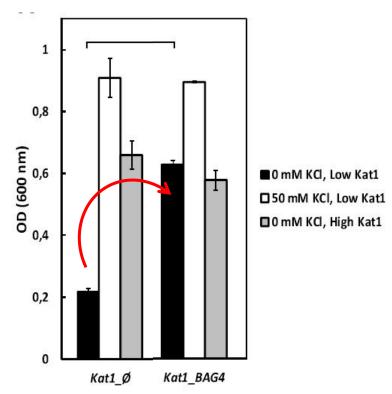
Collaboration with Lynne Yenush (University of Valencia, Spain)

Electrophysiology

Split-Ubiquitin protein-protein interaction screen searching for KAT1 interactors

BAG4 protein was found to physically interact with KAT1 in yeast and also increase K+ uptake.



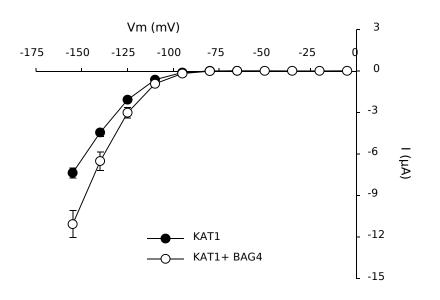


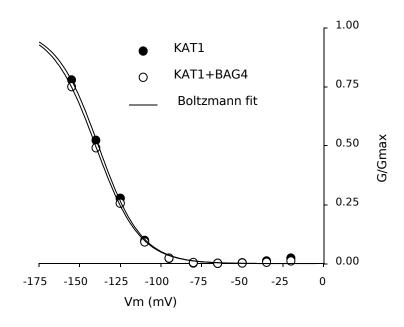
Looking for confirmation of BAG4 effect on KAT1 activity in oocyte?



Collaboration with Lynne Yenush (University of Valencia, Spain)

Electrophysiology





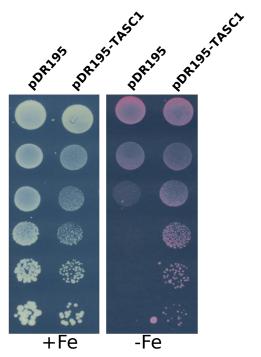
BAG4 co-expression increases **KAT1** current in *Xenopus* oocyte

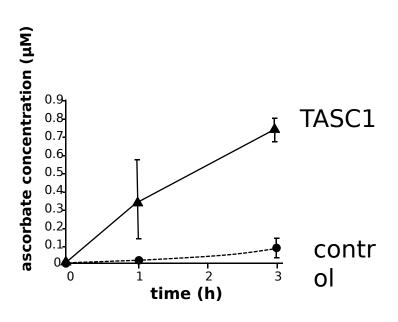
- No shift in KAT1 voltage-dependence was observed under coexpression
- Consistent with higher accumulation of active KAT1 channels at the oocyte cell surface upon co-expression with BAG4

Tracer uptake/efflux

Yeast screen to identify a transporter for Ascorbate efflux: TASC1

TASC1 increases Ascorbate efflux in yeast





Looking for confirmation of Ascorbate efflux in oocyte?

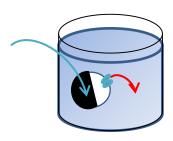


Collaboration with Diego Almeida and Stéphane Mari (TSF)

Tracer uptake/efflux

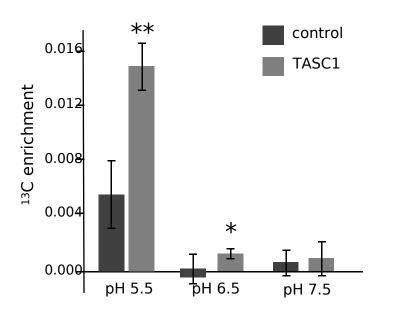


¹³[C] Ascorbate



J0 : Injection of TASC1 cRNA

J2-J3: Injection of ¹³[C] Ascorbate and Efflux assay



¹³[C] Mass spectrometry (P. Tillard)

TASC1 expression in oocyte increases Ascorbate efflux at acidic pH





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E2M : Ecole d'Electrophysiologie de Montpellier

Theorical training in March
Practical training in May: Antony Chung-You-Chong (IGF)



26-29 Novembre : 6 Master students Also for PhD student





MASTER 2
Biologie Fonctionnelle des Plantes
Module ED GAIA

MISTRAL international school on ion and water transport in plants

W4: Voltage-clamp analyses of transporters and voltagegated channels ("whole-cell" current recordings), as examples of non-rectifying and rectifying conductances (Martin, Claire, Nonor): 6 attendees

W5: Patch-clamp analysis of a voltage-gated channel: single channel recording (Martin, Claire, Nonor): 3 attendees





Welcoming and training of young foreign researchers for short courses:

* Limin Wang (Cambridge, UK) : Membrane potential recordings on roots

And as usual for expression and characterization of ion channels in Xenopus oocytes :

* Kumkum Kumari (India)





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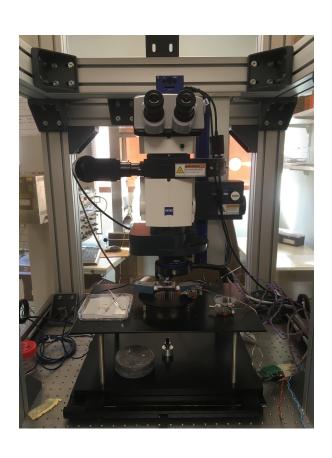
Facilities: Certification from veterinary services, laser setup improvement,

2. News for 2019 and development





Development of in planta voltage-clamp setup (new digidata/software)



JB Thibaud

Motorized stereo (x320) and mono (x1000) microscope

For recordings of intracellular membrane potential and ionic concentrations

Purchase of digidata and acquisition software in December 2018

Switch from Axon/pClamp to ADInstrument Under discussion



Coming of the new team of Alexis de Angeli «Coordination des flux ioniques et signalisation dans les cellules végétales»

Improvement of our Patch-clamp setups



Poste Single channel



Poste Imagerie





Electrophysiology and heterologous expression

New rules for xenopus users because of the certification of the lab



For Xenopus surgery: training qualification needed (niveau 2)

≠ Will be collectively discussed among users



So far, wideopen service delivery in heterologous expression and electrophysiological experiments: https://www1.montpellier.inra.fr/ibip/bpmp/Intranet/reservations. htm

Will also be discussed with Christophe and Alain

My wish:

To help users, to train new beginners, to develop a better tool, to increase our access and collaborations: JUST COME AND DISCUSS!