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reproduction in any medium, provided the original author and source are credited.
A new species and new records of Tetranychidae (Acari: Trombidiformes) from Saudi Arabia, with a key to world species of Mixonychus Ryke and Meyer

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Original research

ABSTRACT

The spider mite genus Neotetranychus Trägårdh is reported for the first time from Saudi Arabia (SA) based on N. longisetus n. sp., collected from Heliotropium arbrainense (Boraginaceae). The new species is described and illustrated from the adult female, and three species, Mixonychus (Bakerina) aculus (Chaudhri), Paraplonobia (Anaplonobia) harteni (Meyer) and Sonotetranychus daleae (Tuttle and Baker) (Acari: Tetranychidae) are reported as new to the mite fauna of SA. Mixonychus (B.) aculus (Chaudhri) is re-described and illustrated based on adult female, collected from an exotic mesquite species from Mexico and Caribbean, Prosopis juliflora (Sw.) DC. (Fabaceae). Moreover, the species, Evertella orissaensis (Prasad) n. comb., is moved from the genus Mixonychus Ryke and Meyer to the genus Evertella Meyer and Mixonychus (Bakerina) ganjuis (Qian, Yuan and Ma) n. comb., is moved to the subgenus Bakerina Chaudhri from the subgenus Mixonychus. A key to the world species of the genus Mixonychus is also provided. Additionally, legs chaetotaxy of all known species of the genus Mixonychus is given.

Keywords aculus; Bakerina; longisetus; Neotetranychus; re-description; spider mites

Zoobank http://zoobank.org/3302B394-5608-4184-8307-D6F66B951196

Introduction

Mixonychus Ryke and Meyer and Neotetranychus Trägårdh are small spider mite genera consisting of nineteen and ten species, respectively (Migeon and Dorkeld, 2006–2018). The economic importance of these two genera is not known yet.

Ryke and Meyer (1960) erected the genus Mixonychus based on type species Mixonychus acaciae Ryke and Meyer from South Africa. The genus Mixonychus is morphologically close to Brevinychus Meyer and Evertella Meyer. It can be differentiated from Brevinychus by longer empodial claws as compared to the pads of the true claw, whereas in the later empodial claws are short, and about as long as the pads of true claws. Mixonychus is different from Evertella by having 10 pairs of dorsal body setae as compared to nine pairs setae in latter (Meyer, 1987; Bolland et al., 1998).

Chaudhri (1971) erected the genus Bakerina with the following characters states; the dorsal propodosoma medially with reticulations and hysterosoma with broken and irregular striations. Gutierrez (1985) treated Bakerina as a subgenus of genus Mixonychus and divided Mixonychus into three subgenera on the basis of dorsal integument ornamentation, namely Mixonychus Ryke and Meyer, Bakerina Chaudhri and Tylochus Miller. Species in the subgenus Mixonychus have a dorsum with protuberances that form a reticulate pattern (four species); species of the subgenus Tylochus have a striate dorsum with spinules, and lacking...
protuberances (four species); and species of the subgenus Bakerina have a dorsum with simple striations, lacking protuberances and spinules (eleven species) (Bolland et al., 1998; Migeon and Dorkeld, 2006–2018). Most of the species belonging to the genus Mixonychus have been reported from Asia, Africa, Australia and the Americas. Recently, Mixonychus (B.) murrayae (Gao and Ma), M. (B.) nanchangensis (Ma and Yuan) and M. (B.) ganjuis (Qian, Yuan and Ma), from China, were re-described and illustrated (Yi and Jin, 2017). To date, there is no key for the species of Mixonychus in the world.

The genus Neotetranychus is characterized by having the empodium split distally, long dorsal body setae set on tubercles, cuticle of the female opisthosoma with varying morphology including patterns of strong striae or clusters of stellate or reticulate protuberances, female with two pairs of pseudoanal setae (ps1,2) (Meyer, 1987; Bolland et al., 1998). Not including the new species proposed here, the genus is comprised of ten nominal species, five of them reported from Central or South America (N. peniculus Aranda, N. raphidoseta Aranda, N. granifer Feres and Flechtmann, N. asper Feres and Flechtmann, N. gloriosus Estebanes and Baker), two from Europe (the type species N. rubi Tragardh, and N. rubicola Bagdasarian), one species each from Africa (N. decorus Meyer and Bolland), Australia (N. victoriae Davis) and Thailand (N. lek Flechtmann). A key to the all previously known species of Neotetranychus was provided by Flechtmann (2013).

Thirty tetranychid mite species belonging to twelve genera have been reported previously from SA (Alatawi and Kamran, 2018; Kamran et al., 2018). In the present study two genera; Mixonychus (Bakerina Chaudhri), and Neotetranychus Trägårdh, also three species M. (B.) aculus (Chaudhri), Paraplonobia (Anaplonobia) harteni (Meyer) and Sonotetranychus daleae (Tuttle and Baker) are recorded for the first time from SA. Neotetranychus longisetus n. sp., is described and illustrated based on adult females collected from a native species of Heliotropium arbinense (Boraginaceae). In addition, Mixonychus (B.) aculus (Chaudhri), is re-described and illustrated based on specimens collected from an exotic mesquite species (native to Mexico, South America and the Caribbean), Prosopis juliflora (Sw.) DC. (Fabaceae). Everterella orissaensis (Prasad) is n. comb., transferred from Mixonychus to the genus Everterella Meyer as opisthosomal setae f1 absent, and Mixonychus (B.) ganjuis (Qian, Yuan and Ma) n. comb., is moved to the subgenus Bakerina (from subgenus Mixonychus) as it lacks clusters of protuberances on the dorsal integument. A key and details of the legs chaetotaxy, is provided for the world species of the genus Mixonychus.

Materials and methods

Spider mites were collected from a diversity of host plants in different localities across Saudi Arabia during 2016–2018. The mite specimens were collected by shaking the aerial parts over a piece of white paper. The mites moving on paper were collected with the help of camel hair brush and preserved in small vials containing 70% ethanol, then mounted in Hoyer’s medium under a stereomicroscope (SZX10, Olympus®, Tokyo, Japan). The specimens were examined and identified under a phase contrast microscope (BX51, Olympus®, Japan) using diagnostic keys and available literature. Different mite body parts were imaged using an auto-montage software system (Syncroscopy, Cambridge, UK) attached with phase contrast microscope (DM2500, Leica®, Germany) and then drawn with Adobe Illustrator (Adobe System Inc., San Jose, CA, USA). All measurements are given in micrometers (μm). The morphological terminology used in this study followed Lindquist (1985). Legs setal counts are presented as the number of tactile setae followed by the number of solenidia and eupathidia in parentheses. The body measurements are presented for the holotype followed by the range for the paratypes in parentheses when types were examined. All collected specimens, including type specimens of the new species have been deposited at King Saud University Museum of Arthropods (KSMA, Acarology section), Department of Plant Protection, College of Food and Agriculture Sciences, King Saud University, Riyadh, Saudi Arabia.
Results

Family Tetranychidae Donnadieu
Subfamily Tetranychinae Berlese
Tribe Tetranychini Reck
Genus Neotetranychus Trägårdh

Type species: Neotetranychus rubi Trägårdh, 1915: 33.

Diagnosis — Based on the Pritchard and Baker (1955) and Meyer, 1987.

Female: Dorsal body setae long and set on strong tubercles; opisthosomal integument mostly with transverse striae, wrinkled or reticulated clusters; legs empodia curved, uncinate and undivided along most of its length, with distal tip divided into pairs of fine hairs; two pairs of pseudoanal setae (ps$_{1-2}$).

Neotetranychus longisetus n. sp.

(Figures 1–4)

Zoobank: 553E8F35-7EF4-46F7-B567-DB5CF26F0093

Diagnosis (Based on adult female) — Propodosoma medially with longitudinal striae; hysterosoma medially with transverse striae and laterally with longitudinal to oblique striae; striae encircling bases of each dorsal seta, forming concentric rings; cuticle with rows of regular, shallow, scallop-shaped projections; dorsal body setae acicular, of even thickness along length, finely serrated, inserted on distinct tubercle; most of the dorsal setae twice as long as...
the distance to base of setae next consecutive row, dorsocentral setal pairs cl, dl, el and fl subequal distance apart.

**Description of the Adult female (n = 12)**

Length of idiosoma (excluding gnathosoma) 331 (315–345), (including gnathosoma) 404 (390–420), maximum width at the level of row c setae 263 (255–270).

**Dorsum** (Figures 1A-C) — Dorsal integument entirely striated, striae longitudinal on propodosoma, striae transverse medially and longitudinal to oblique laterally on hysterosoma; cuticle with rows of regular, shallow, scallop-shaped projections (Figures 1A-B); dorsum with 13 pairs of setae; seta c3 inserted at lateral margin of body, often situated ventrally; setae v2, sc2, c3 and h1 shorter than other dorsal setae, almost equal in length; dorsocentral setal pairs cl, dl, el and fl subequal distance apart. Length of dorsal setae: v2 63 (60–65), cl 110 (107–114), sc2 63 (60–65), e1 142 (135–148), c2 126 (120–130), cl 65 (62–68), dl 142 (138–150), d2 129 (120–135), e1 89 (84–93), e2 123 (120–128), fl 105 (100–120), f2 84 (80–90), h1 63 (60–68); distance between dorsal setae: v2−v2 53 (50–55), cl−cl 110 (107–114), sc2−sc2 205 (200–210), cl−c1 42 (39–43), c2−c2 126 (120–135), c3−c3 263 (250–280), dl−dl 42 (39–45), d2−d2 147 (140–155), e1−el 37 (35–42), e2−e2 131 (125–140), fl−fl 37 (35–40),

**Figure 2** *Neotetranychus longisetus* n. sp., adult female: A – Venter; B – Genital area. Scalebar A, B = 100 μm.
Venter (Figures 2A-B) — Ventral idiosoma with transverse striations from setae 1a to just anterior to ag, pregenital area with transverse to arching striation (Figure 2B); striations on venter without lobes. Length of ventral setae: 1a 53 (51–55), 3a 55 (52–58), 4a 53 (50–55), 1b 63 (60–65), 1e 53 (51–56), 2b 42 (39–44), 3b 50 (50–52), 4b 53 (50–55); distance between inter-coxal and coxa setae: 1a–1a 26 (25–28), 1b–1c 11 (10–12), 3a–3a 53 (50–55), 4a–4a 63 (60–65); aggenital setae: ag 55 (53–57), ag–ag 62 (60–65); two pairs of genital setae: gl 27 (25–29), g2 32 (31–34), gl–g1 26 (23–28), g2–g2 70 (65–74); two pairs of pseudoanal setae: ps1 16 (15–18), ps2 15 (13–16), ps1–ps2 6 (5–7), ps1–ps1 32 (30–35), ps2–ps2 21 (20–23).

Gnathosoma (Figure 3) — Ventral infracapitular setae m 50 (48–53), m–m 35 (33–37). Palp femur and genu each with one setae, d 28 (26–29), l” 18 (16–19); palp tibia with three setae, d 22 (19–22), l” 16 (15–18), l’ 15 (15–16) and a palp tibial claw; palp tarsus 14 long, 11 wide, with three simple setae a 10 (9–11), b 7 (7–9), c 8 (7–10), three eupathidia suζ 7 (6), ul”ζ = ul’ζ 6 (5–7), a solenidion ω 5 long; stylophore with weak anterior notch; peritremes terminating in short hook, like a golf-club (Figure 1C); terminal sensillum of palp tarsus conical, long, pointed about three times as long as wide (Figure 3).


Male and immature stages Unknown.

Figure 3 Neotetranychus longisetus n. sp., adult female: Palp. Scalebar = 50 μm.
**Etymology** — The specific epithet refers to the length of dorsocentral setae which are comparatively longer than that of other species.

**Material examined** — Holotype female and 11 paratypes females, *Heliotropium arbainense* (Boraginaceae), Taif, 21°21.151’N, 040°19.515’E, elevation 1843 m, 11 September 2017, coll. E. M. Khan and M. U. Rehman. One Holotype female (KSMA-ARL-19/5) and 11 paratypes female (KSMA-ARL-19/6-16) has been deposited at King Saud University Museum of Arthropods (KSMA, Acarology section), Department of Plant Protection, College of Food and Agriculture Sciences, King Saud University, Riyadh, Saudi Arabia.

**Remarks** — *Neotetranychus longisetus* n. sp., is different from all other species of *Neotetranychus* by entire dorsum striated and with scallop-shaped structures. However, the new species resembles *N. lek* Flechtmann 2013. *Neotetranychus lek* has longitudinal striae between opisthosomal setae e1−e1 vs. transverse striae on *N. longisetus* n. sp., femur and genu.
III each 4 setae vs. 4 and 3 on *N. longisetus* and femur and genu IV each with 3 setae vs. 4 and 3 setae on *N. longisetus* n. sp..

**New records for Saudi Arabia**

*Genus Mixonychus* Ryke and Meyer

*Mixonychus (Bakerina) aculus* (Chaudhri)


Re-description of adult female (*n* = 6)

(Figures 5–8)


**Gnathosoma** (Figure 7) — Ventral infracapitular setae *m* 35–39, *m*–*m* 19–21. Palp femur and genu each with one seta, *d* 32–35, *l* 23–25; palp tibia with three setae, *d* 20–22, *l* 17–19, *l*′ 15–16 and a palp tibial claw; palp tarsus 14 long, 11 wide, with three simple setae *a* 8–10, *b* 9–10, *c* 10–12, three eupathidia *su*ζ 5–6, *ul*ζ = *ul*ζ 7–9, a solenidion *ω* 11 long; peritremes terminating in hooked; terminal sensillum of palp tarsus conical, long, pointed about 3 time long as wide to the base.


**Male and immature stages** Unknown.

**Material examined** — Four females, *Prosopis juliflora* (Fabaceae) (native to Mexico, South America, and the Caribbean), Alhasa 27°55.0143′N, 047°66.8970′E, 21 April 2018,
Figure 5 Mixonychus (Bakerina) aculus (Chaudhri), adult female: A, B – Dorsum; C – Dorsal seta. Scalebar A, B = 100 μm.

coll. M. Kamran, J. H. Mirza and H. M. Saqib. Four females (KSMA-ARL-19/16-20) have been deposited at King Saud University Museum of Arthropods (KSMA, Acarology section), Department of Plant Protection, College of Food and Agriculture Sciences, King Saud University, Riyadh, Saudi Arabia.

**Distribution** — Pakistan (Chaudhri, 1971) and SA in present study.

**Known hosts** — *Senegalia modesta* (syn. *Acacia modesta*, Mimosaceae) (native to India, Pakistan and Afghanistan), *Albizia lebbeck* (paratypes host, Mimosaceae) (native to Indomalay region, Papua New Guinea and northern Australia), *Platycladus orientalis* (Cupressaceae) (native to Korea and northern China), *Salvadora oleoides* (Salvadoraceae) (paratypes host; native to India, Pakistan and southern Iran), *Vachellia nilotica* (syn. *Acacia arabica*) (Mimosaceae) holotype host (native to Africa, Middle East and India) (Chaudhri, 1971; Chaudhri *et al.*, 1974).

**Remarks** — We could not find the type specimens of *M. (B.) aculus* from the depository. The specimens of *M. (B.) aculus* collected from Saudi Arabia are morphologically similar to the original description except minor differences in length of dorsal and ventral body setae; $v_2$
38–43 vs. 42, sc1 49–53 vs. 57 sc2 36–42 vs. 47, cl 37–46 vs. 44, c2 41–47 vs. 47 dl 40–46 vs. 52, d2 44–50 vs. 49, el 44–49 vs. 52, e2 49–53 vs. 49, fl 47–52 vs. 52, f2 38–41 vs. 47, hl 44–48 vs. 49, l1 43–47 vs. 39, l2 41–44 vs. 42, l3 31–34 vs. 36 and solenidia on both tarsi III and IV 10–12 vs. 16 in original description.

**Genus Sonotetranychus Tuttle, Baker and Abbiatiello**

*Sonotetranychus daleae* (Tuttle and Baker)


*Sonotetranychus daleae* (Tuttle and Baker), Tuttle, Baker and Abbiatiello, 1976: 70.

**Material examined** — Two females, *Fachellia nilotica*, Taif, 21°30.112’N, 040°29.118’E,
Mixonychus (Bakerina) aculus (Chaudhri), adult female: Palp. Scalebar = 50 μm.

- **Figure 7**

**Material examined** — Four females, *Indigofera spinosa* (Leguminosae) (native to the region and northeast Africa), Taif, 21°33.926’N, 040°07.260’E, elevation 643 m, 15 October 2016, coll. M. U. Rehman and M. Kamran. Four females (KSMA-ARL-19/29-33) have been deposited at King Saud University Museum of Arthropods (KSMA, Acarology section), Department of Plant Protection, College of Food and Agriculture Sciences, King Saud University, Riyadh, Saudi Arabia.

**Distribution** — Yemen (Meyer, 1996).

**Known hosts** — Unknown plant species (Meyer, 1996).
New combination

Evertella orissaensis (Prasad, 1975), n. comb.

Mixonychus (Bakerina) orissaensis (Prasad): Bolland et al., 1998: 86.

Distribution — India.

Known hosts — Allium sp., (Amaryllidaceae), Artocarpus heterophyllus (Moraceae) (Prasad, 1975).

Remarks — Evertella was erected by Meyer (1987) to accommodate Mixonychus (Mixonychus) koegasensis (Meyer, 1974). Like Mixonychus (Mixonychus), Evertella is characterized by having the dorsal integument finely striate and with protuberances that produce a reticulate appearance, and a simple claw-like empodium. Evertella was separated from Mixonychus...
because of dorsal opisthosomal setae f1 are absent (present in Mixonychus).

Prasad 1975 described the dorsal integument of Bakerina orissaensis as striate with comparatively large, round punctations, and Bolland et al. (1998) placed B. orissaensis within Mixonychus (Bakerina) due to the striate dorsum and overlooked the absence of seta f1 in B. orissaensis which distinguish Evertella from Mixonychus. Therefore, due to absence of f1 seta, Mixonychus (Bakerina) orissaensis is transferred from Mixonychus (Bakerina) to Evertella.

Mixonychus (Bakerina) ganjuis (Qian, Yuan and Ma, 1980), n. comb.

Mixonychus (Mixonychus) ganjuis Qian, Yuan and Ma, 1980: 80.
Mixonychus (Mixonychus) ganjuis Qian, Yuan and Ma; Yi and Jin, 2017: 1296–1301.

**Distribution** — China.

**Known hosts** — Amelanchier sp., (Rosaceae), Citrus reticulata (Rutaceae) (Yi and Jin 2017).

**Remarks** — The species M. (B.) ganjuis having dorsal integument striated, without lumps and spinules, which was misplaced previously in the subgenus Mixonychus (Yi and Jin, 2017), is transferred from subgenus Mixonychus to subgenus Bakerina.

### Table 1 Legs chaetotaxy of world species of the genus Mixonychus based on literature (tarsal setal counts are presented as total number of tactile setae and eupathidia (total number of solenidia + number of duplex pairs)).

<table>
<thead>
<tr>
<th>Species</th>
<th>Femora</th>
<th>Genua</th>
<th>Tibiae</th>
<th>Tarsi</th>
<th>Reference</th>
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<tr>
<td>acaciae</td>
<td>8−6−3−2</td>
<td>5−5−3−3</td>
<td>(1or 3φ)−5−5−5</td>
<td>12(1or 2ω+2dup.)−10(1ω+1dup.)−9(1ω)−9(1ω)</td>
<td>Meyer, 1974</td>
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<tr>
<td>neoacaciae</td>
<td>7−5−3−2</td>
<td>5−5−3−3</td>
<td>8(1φ)−5−5−5</td>
<td>9(1ω)−9(1ω)−9−9</td>
<td>Smiley &amp; Baker, 1995</td>
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<tr>
<td>dulcis</td>
<td>8−6−3−2</td>
<td>5−5−3−3</td>
<td>8 or 9(1φ)−5−5−5</td>
<td>12(1ω+2dup.)10(1ω+1dup.)−9(1ω)−9(1ω)</td>
<td>Meyer, 1974</td>
</tr>
<tr>
<td>cruzae</td>
<td>9−8−4−4</td>
<td>5−5−4−4</td>
<td>9(1φ)−8−6−7</td>
<td>11(1ω+2dup.)−10(1ω+1dup.)−5(1ω)?</td>
<td>Corpuz-Raros, 1978</td>
</tr>
<tr>
<td>tasmaniensis</td>
<td>??</td>
<td>6(1φ)−5−4−4</td>
<td>?−7(1ω)−7(1ω)</td>
<td>Miller, 1966</td>
<td></td>
</tr>
<tr>
<td>transvaalensis</td>
<td>6−5−3−2</td>
<td>5−5−3−3</td>
<td>8(1φ)−7−5−6</td>
<td>9(1ω+2dup.)−7(1ω)−7(1ω)</td>
<td>Meyer, 1974</td>
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<tr>
<td>jayaraji</td>
<td>7−7−4−4</td>
<td>5−5−3−3</td>
<td>7−7−6</td>
<td>11(2dup.)−9(1ω)−7−7</td>
<td>Karuppachamy &amp; Mohanasundaram, 1987</td>
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</table>

**Mixonychus (Tylonychus)**

<table>
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<th>Species</th>
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<th>Tibiae</th>
<th>Tarsi</th>
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<td>aculus</td>
<td>8−6−4−4</td>
<td>5−5−3−3</td>
<td>10−7−6−7</td>
<td>*15−12−9−9</td>
<td>Chaudhri, 1971</td>
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<tr>
<td>aestiva</td>
<td>7−6−4−4</td>
<td>5−5−4−3</td>
<td>9(1φ)−8(1φ)−5−7</td>
<td>10(1ω+2dup.)−10−9−9</td>
<td>Present Study</td>
</tr>
<tr>
<td>carracios</td>
<td>8−6−4−4</td>
<td>5−5−3−3</td>
<td>9(1φ)−7−6−7</td>
<td>*14−12−10−10</td>
<td>Chaudhri, 1971</td>
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<td>citaceus</td>
<td>7 or 6−6−3−2</td>
<td>5 or 4−5−3−3</td>
<td>7 or 6(1φ)−6 or 5(1φ)−5−5</td>
<td>9(1ω+2dup.)−9 or 10(1ω+1dup.)−7(1ω)−7(1ω)</td>
<td>Flechtmann &amp; Mesa, 2016</td>
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<tr>
<td>ganjuis</td>
<td>9−7−4−4</td>
<td>5−5−4−4</td>
<td>9(1φ)−8−6−7</td>
<td>14(1ω+2dup.)−11(1ω+1dup.)−9(1ω)−9(1ω)</td>
<td>Yi &amp; Jin, 2017</td>
</tr>
<tr>
<td>lepidus</td>
<td>8−6−4−4</td>
<td>5−5−3−3</td>
<td>10−7−6−7</td>
<td>*15−12−9−9</td>
<td>Chaudhri, 1971</td>
</tr>
<tr>
<td>murrayae</td>
<td>9−7−4−4</td>
<td>5−5−4−3</td>
<td>9(1φ)−8−6−7</td>
<td>14(1ω+2dup.)−11(1ω+1dup.)−9(1ω)−9(1ω)</td>
<td>Yi &amp; Jin, 2017</td>
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<td>nanchangensis</td>
<td>9−7−4−4</td>
<td>5−5−4−4</td>
<td>9(1φ)−8−6−7</td>
<td>14(1ω+2dup.)−11(1ω+1dup.)−9(1ω)−9(1ω)</td>
<td>Yi &amp; Jin, 2017</td>
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<td>normalis</td>
<td>8−6−4−4</td>
<td>5−5−3−3</td>
<td>9(1φ)−7−6−7</td>
<td>*14−12−10−10</td>
<td>Chaudhri, 1971</td>
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<tr>
<td>thailandicus</td>
<td>9 or 7−7−4−4</td>
<td>5−5−4 or 3−3</td>
<td>9(1φ)−8−6−6</td>
<td>9 or 8(1ω)−7(1ω)−9(1ω)−9(1ω)</td>
<td>Tangkansasing, 1988</td>
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<tr>
<td>zolanzensis</td>
<td>8−7−5−4</td>
<td>5−5−4−3</td>
<td>8(1φ)−8−6−7</td>
<td>8(1ω+2dup.)−8(1ω+1dup.)−9(1ω)−8(1ω)</td>
<td>Lo &amp; Ho, 1989</td>
</tr>
</tbody>
</table>

* Total setae on tarsi with duplex setae and solenidia as mentioned in description.
? No information found in relative description.
Key to world species of the genus Mixonychus Ryke and Meyer (Based on adult female)

1. Dorsal integument with small protuberances forming a reticulate pattern or producing the appearance of a reticulate pattern ................ subgenus Mixonychus Ryke and Meyer – 2
   — Dorsal integument without protuberances; integument with/without spinules present ...... 4

2. Length of dorsal body setae less than half the longitudinal distance to the setae in the next row; only dorsal opisthosomal setae $f_2$ and $h_1$ set on tubercles .......................................................... $M. (M.)$ dulcis Meyer, South Africa
   — Length of dorsal body setae more than half the longitudinal distance to the setae in the next row; almost all dorsal opisthosomal setae set on tubercles .............................................. 3

3. Femora I and II with 8 and 6 setae, respectively................................................................. $M. (M.)$ acaciae Ryke and Meyer, South Africa
   — Femora I and II with 7 and 5 setae, respectively......................................................... $M. (M.)$ neoacaciae Smiley and Baker, Yemen

4. Dorsal integument with spinules ............................................................... subgenus Tylonychus Miller – 5
   — Dorsal integument without spinule........................................................... subgenus Bakerina Chaudhri – 8

5. Dorsal opisthosomal setae long, clearly longer than the bases of the setae in the next row..... 6
   — Dorsal opisthosomal setae short, not reaching the bases of the setae in the next row...... 7

6. Dorsal body setae set on strong tubercles; female propodosoma integument with simple striations; dorsal opisthosomal setae $f_2$ short, as compared to other dorsal setae ................ $M. (T.)$ tasmaniensis (Miller), Australia
   — Dorsal body setae not set on tubercles; female propodosoma integument with a reticulate pattern; dorsal opisthosomal setae $f_2$ as long as other dorsal body setae .............................................. $M. (T.)$ jayaraji (Karuppuchamy and Mohanasundaram), India

7. Dorsal body setae stout, rod shaped; genua III and IV with 3 setae; femora I–IV with 6–5–3–2 setae ............................................................... $M. (T.)$ transvaalensis (Meyer), South Africa
   — Dorsal body setae oblong, lanceolate; genua III and IV with 4 setae; femora I–IV with 9–8–4–4 setae ............................................................... $M. (T.)$ cruzae (Corpuz-Raros), The Philippines

8. Dorsal opisthosomal setae $d_1$, $e_1$ as long as or longer than the distance to the bases of setae in the next row ........................................................................................................ 9
   — Dorsal opisthosomal setae $d_1$, $e_1$ obviously shorter than the distance to the bases of the setae in the next row (approx. 2/3 the distance) ................................................................. 14

9. Femora I and II with 9 and 7 setae, respectively; genua III and IV each with 4 setae ...... 10
   — Femora I and II with 8 and 6 setae, respectively; genua III and IV each with 3 setae .... 11

10. Medial integument of propodosoma striated; dorsal setae gradually expanded distally (weakly clavate) .................................................. $M. (B.)$ ganjuis (Qian, Yuan and Ma), China
    — Medial integument of propodosoma reticulated; dorsal setae not expanded distally, of even thickness along length .......................................................... $M. (B.)$ nanchangensis (Ma and Yuan), China

11. Dorsal body setae lanceolate; tibia I without lanceolate seta; tarsi III and IV with long solenidion (16 μm long) ................................................................. 12
    — Dorsal body setae of even thickness along length, weakly to not tapering, with broadly blunt tips; tibia I with 3 lanceolate setae; tarsi III and IV with short solenidion (5 μm long) .... 13
12. Dorsal opisthosomal setae c1 (44 μm) shorter than the distance c1–d1 ............................................. M. (B.) aculus (Chaudhri), Pakistan
— Dorsal opisthosomal setae c1 (60 μm) reaching the base of setae d1 ............................................. M. (B.) lepidus (Chaudhri), Pakistan

13. Peritremes ending in simple unexpanded bulb; dorsal opisthosomal f1 70 μm long and f2 44 μm long ............................................. M. (B.) carracis (Chaudhri), Pakistan
— Peritremes ending in an expanded bulb; dorsal opisthosomal setae f1 83 μm long and f2 47 μm long ............................................. M. (B.) normalis (Chaudhri), Pakistan

14. Dorsal body setae gradually expanding along length, elongate obovate; femur III with 3 or 4 setae ............................................. 15
— Dorsal body setae of even thickness along length; femur III with 5 setae ............................................. M. (B.) ziolanensis (Lo and Ho), Taiwan

15. Femur II with 6 setae; tibia III with 5 setae ............................................. 16
— Femur II with 7 setae; tibia III with 6 setae ............................................. 17

16. Femur III with 3 setae; tibia II with 5 setae ............................................. M. (B.) citraeus Flechtmann and Mesa, Colombia
— Femur III with 4 setae; tibia II with 8 setae ............................................. M. (B.) aestival (Tseng), Taiwan

17. Tibia IV with 7 setae ............................................. M. (B.) murrayae (Gao and Ma), China
— Tibia IV with 6 setae ............................................. M. (B.) thailandicus (Tangkanasing), Thailand

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References


