THE GENUS *ORNITHONYSSUS* SAMBON 1928 IN THE ETHIOPIAN REGION: DESCRIPTION OF A NEW SPECIES AND A REDESCRIPTION OF *O. ROSEINNESI* (ZUMPT & TILL, 1953) (ACARINA, MESOSTIGMATA)

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HABITAT
IN ETHIOPIAN
REGION
ACARI
OF MEDICAL
AND VETERINARY
IMPORTANCE

ABSTRACT: A new macronyssid mite is described based on material collected by the authors in July 1981 from an elephant shrew (*Elephantulus myurus* Thomas and Schwann, 1906) at Kimberley, South Africa. The female of *Ornithonyssus roseinnesi* (Zumpt and Till, 1953) is redescribed and a key given to females of the Ethiopian species of *Ornithonyssus*. New host records for the genus in the Ethiopian region are presented.

WIRTE IM ÄTHIOPISCHE RAUM KRANKENMILBEN ZUSAMMENFASSUNG: Auf der Grundlage von Material welches die Autoren im Juli 1981 von der Elephantenschrew (*Elephantulus myurus* Thomas und Schwann, 1906) in Kimberley, Südafrika, gesammelt haben, wird hier ein neue Milbe der Familie *Macronyssidae* beschrieben. Ausserdem wird das Weibchen der Art *Ornithonyssus roseinnesi* (Zumpt and Till, 1953), neu beschrieben und ein Schlüssel zur Identifizierung der äthiopischen Ornithonyssinen gegeben. Neue Wirte für diese Gattung im äthiopische Raum werden aufgeführt.

Introduction

Recently the genus *Ornithonyssus* has been reviewed on a worldwide basis by MICHERDZINSKI (1980) but apart from the description of a new species, all his information on African species is based on the work of ZUMPT (1961). Since the publication of ZUMPT's book new records of *Ornithonyssus* mites have come to light from African mammals and birds, including the discovery of a new species.

In July 1976 a single female mite was taken from a Namaqua rock mouse Aethomys namaquensis (A. Smith, 1834) captured at Onseepkans, Kenhardt district, Cape Province. This mite resembled Ornithonyssus roseinnesi (Zumpt and Till, 1953) in general appearance but had setae present on the opisthonotal region of the dorsal shield (behind seta J4) — a feature absent in the figures of ZUMPT and TILL (1953) and MICHERD-ZINSKI (1980). On examination of the type series at the South African Institute for Medical

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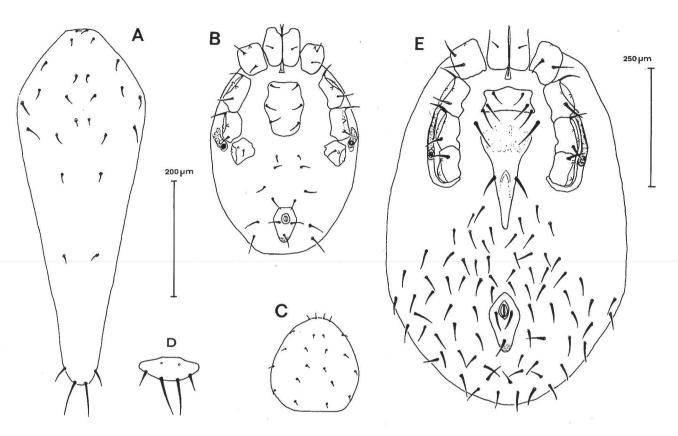


Fig. 1: Ornithonyssus capensis n. sp.

A. — Female, dorsal shield; B, C, D. — Protonymph, venter (B), podonotum (C), pygidium (D); E. — Female holotype, venter.

Research, Johannesburg, the original descriptions were found to be inadequate. In common with our specimen the type material was seen to have setae present posterior to seta J4. It was decided that they were the same species and that a redescription of O. roseinnesi should be made.

The redescription of *O. roseinnesi* together with a description of the new species and a key to the females of the Ethiopian species of *Ornithonyssus* are presented in this paper. The system of setal nomenclature proposed by MICHERDZINSKI (1980) is used for the terminology of the setae of the opisthonotum, with the exception of the I series which is replaced by the capital *J*. New host records for the genus in Africa are also given.

Ornithonyssus capensis n. sp. (Fig. 1, A-E)

Diagnosis.

The female of this species is easily recognised by having only two pairs of relatively long setae of the dorsal shield posterior to seta J4, by an extremely short and well defined peritreme, and by the chaetotaxy of genu IV, $2\frac{2}{1}\frac{3}{1}2$ (11). The protonymph has a podonotum with 11 pairs of very short setae, a pygidium with two pairs of setae and genu IV with chaetotactic formula $1\frac{2}{0}\frac{2}{0}1$ (6).

Female

Idiosoma (holotype) with a greatest width of 560 μm, length 770 μm. Dorsal shield reticulate, long and tapering (605 μ m long, 234 μ m wide), bearing 14 pairs of setae, all short except Z5 and J6 (30 μ m and 55 μ m long respectively), seta i.3 absent. Sternal shield 125 µm wide, 50 µm long posterior margin invaginated almost to the level of seta st 2, st 3 situated on margin of most posterior extension of the plate. Genitoventral shield 72 µm wide at level of genitoventral setae, reticulate, short, tapering sharply to a blunt point posteriorly. Anal shield 140 μm long, 55 μm wide, elongate, broadly rounded anteriorly and tapering to a blunt point posteriorly. Anal setae 30 µm long; adanal setae subequal. Endopodal plates absent. Peritreme well defined, thick and strikingly short barely reaching to posterior margin of coxa II.

Gnathosoma with four pairs of short, simple, slender setae; hypostomal seta III 20 µm long.

Table 1. Segmental chaetotaxy of female and protonymph Ornithonyssus capensis n. sp.

Leg	Coxa	Trochan- ter	Femur	Genu	Tibia	Tarụs
	Female					
I	$0\frac{00}{11}0(2)$	1 2 1 (6)	$2\frac{32}{13}2(13)$	$2\frac{33}{21}2(13)$	$2\frac{33}{21}2(13)$	_
II	$0\frac{0.0}{1.1}0(2)$	$1\frac{00}{21}1(5)$	$2\frac{32}{12}1(11)$	$2\frac{3.2}{11}$ 2(11)	$2\frac{22}{11}$ 2(10)	$3\frac{3}{2}\frac{13}{1-2}3(18)$
III	$0\frac{00}{11}$ 0(2)	$1\frac{10}{21}0(5)$	$1\frac{21}{10}1$ (6)	$2\frac{22}{11}$ 2(10)	$2\frac{12}{11}$ 2 (9)	as II
IV	$0\frac{00}{10}$ 0(1)	$1\frac{10}{21}0(5)$	$0\frac{21}{20}1$ (6)	$2\frac{23}{11}2(11)$	$2\frac{13}{11}$ 2(10)	as II
Protonymph						
I	0 0 0 0 (2)	1 00 1 (4)	2 ²² / ₁₁ 2(10)	1 22 1 (8)	$1\frac{22}{11}$ 1 (8)	_
II	$0\frac{00}{11}0(2)$	$1\frac{00}{02}1(4)$	$1\frac{22}{11}$ 1 (8)	$1\frac{22}{00}1$ (6)	$1\frac{12}{11}$ 1 (7)	$3\frac{313}{202}3(17)$
Ш	$0\frac{00}{11}0(2)$	$1\frac{10}{11}0(4)$	$1\frac{21}{10}1$ (5)	$1\frac{22}{00}$ 1 (6)	$1\frac{12}{11}$ 1 (7)	as II
IV	$0\frac{0}{1}\frac{0}{0}$ 0(1)	$1\frac{1}{2}\frac{0}{0}0(4)$	$0\frac{21}{10}0$ (4)	$1\frac{2}{0}\frac{2}{0}$ 1 (6)	$1\frac{12}{11}$ 1 (7)	as II

Leg measurements : I = 621 μ m, II = 525 μ m, III = 520 μ m, IV = 680 μ m long. Segmental chaetotaxy (Table 1) as for the genus *Ornithonys*-

sus (Micherdzinski, 1980) except genu IV with for- $2\frac{2}{1}\frac{2}{1}$ 3 (11).

■ Protonymph

Idiosoma (average of paratypes) with greatest width 254 μ m, length 345 μ m. Podonotum 159 μ m wide, 152 μ m long, with 11 pairs of short setae 13 μ m long. Pygidium 89 μ m wide, 27 μ m long with two pairs of setae, setae Z5 and J6 27.5 μ m and 41 μ m long respectively. Sternal shield 110 μ m wide, 75 μ m long with three pairs of short sternal setae. Anal shield 35 μ m wide, 55 μ m long. Anal and adanal setae short and subequal. Peritreme short. Gnathosomal setae short, slender.

Leg measurements : I = 322 μ m, II = 269 μ m, III = 267 μ m, IV = 332 μ m long. Segmental chaetotaxy (Table 1) as for genus *Ornithonyssus* except genu IV with formula $1\frac{2}{0}\frac{2}{0}\frac{2}{0}$ 1 (6).

Male — unknown.

Type material

Female holotype, three paratypes and eight protonymphs (coll. A. J. Shepherd, PL 81: 125) Benfontein, Kimberley, Cape Province, South Africa on *Elephantulus myurus* Thomas and Schwann 1906, collected July 1981.

Specimens are in the collection of the South African Institute for Medical Research, Johannesburg.

Ornithonyssus roseinnesi (Zumpt & Till, 1953)

Liponyssus roseinnesi: ZUMPT & TILL, 1953; Ornithonyssus roseinnesi: STRANDTMANN & WHARTON, 1958; Macronyssus roseinnesi: ZUMPT, 1961; Ornithonyssus roseinnesi: MICHERDZINSKI, 1980.

Diagnosis

This species is easily recognised by the extremely short setae over the entire surface of the dorsal shield, by the characteristic shape of the

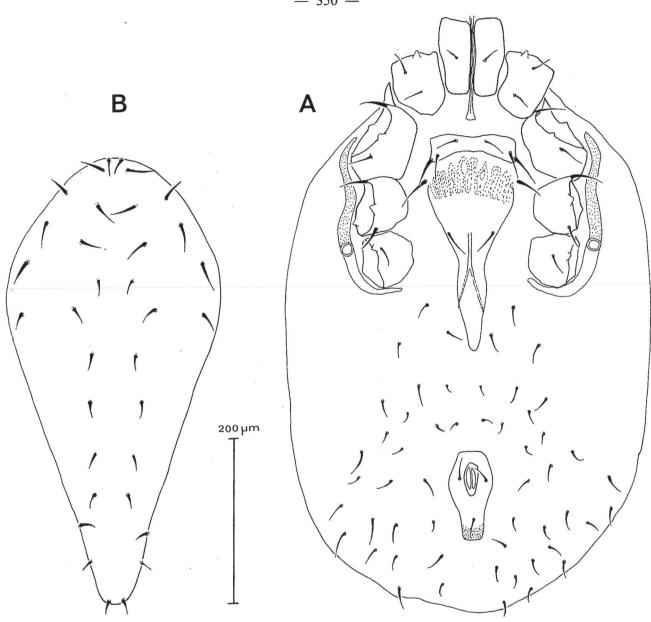


Fig. 2: Ornithonyssus roseinnesi (Zumpt & Till, 1953). A, B. — Female, venter (A), dorsal shield (B).

sternal shield (length/width = 1/5) by the anterodorsal spine present on coxa II.

Female (Holotype no. 1327)

Idiosoma with greatest width 440 μm , 662 μm long. Dorsal shield 262 μm wide, 535 μm long,

elongate pear-shape, tapering posteriorly, slightly reticulate with 17 pairs of short setae (longer anteriorly). Seta I3 absent I6 10 μ m long. Venter of idiosoma with approximately 25 pairs of short (on average 25 μ m long) slender setae. Sternal shield 117 μ m wide, 22.5 μ m long, extremely narrow and much wider than long. Genitoventral

shield tapering to a blunt point posteriorly. Genitoventral setae 60 μ m apart. Anal shield pear-shaped, 60 μ m, wide, 102 μ m long. Anal and adanal setae short, subequal.

Gnathosomal setae simple, slender, of averagelength; hypostomal seta III = $32.5 \mu m$ long. Endopodal plate small, triangular, present opposite coxa II. Peritreme reaches to coxa II.

Leg measurements : I = 462 μ m, II = 370 μ m, III = 335 μ m, IV 425 μ m long. Segmental chaetotaxy as for the genus *Ornithonyssus* (MICHERDZINSKI, 1980). Anterodorsal spine present on coxa II.

- Protonymph Unknown.
- Male Unknown.

Hosts and localities .

The original description was based on 49 females collected from the nest of a rodent, *Rhabdomy's pumilio* (Sparman, 1784), at Goshen, Cape Province. Since then only a single female has been found, taken from *Aethomys namaquensis* (A. Smith, 1834) at Onseepkans, Cape Province in 1976. The great distance between the two collecting sites suggests a wide distribution for this species.

KEY TO THE FEMALES OF THE ETHIOPIAN SPECIES OF Ornithonyssus sambon, 1928

1.	Palpal	trochanter	with	ventral	spine	3
	Palpal	trochanter	witho	out vent	ral spine	2

2. Anterodorsal spur on coxae II present, genus IV with 10 setae, Z5 and J6 small...... roseinnesi (Zumpt & Till, 1953) Anterodorsal spur on coxa II absent, genu IV with 11 setae, Z5 and J6 long..... capensis n. sp. 3. Anterodorsal spur present on coxa II...... 5 Anterodorsal spur absent on coxa II...... 4 4. Three pairs of lateral seta posterior to I4 on the dorsal shield, Z4 and J6 subequal..... sylviarum (Canestrini & Fanzago, 1877) Four pairs of lateral setae posterior to J4 on the dorsal shield, Z5 and J6 subequal, Z4 shorter..... bursa (Berlese, 1888) 5. Dorsal shield narrow, tapering posteriorly..... 6 Dorsal shield otherwise...... 7 6. Anal shield greatly extended posteriorly to form long lobe, large species (idiosoma greater than 1.5 mm), sternal shield with round, pore-like structures. africanus (Zumpt & Till, 1958) Anal shield without lobe, smaller species (idiosoma less than 1 mm), sternal shield without pore-like structures. Cosmopolitan species..... bacoti (Hirst, 1913) 7. Posteroventral spur on coxa III present..... lukoschosi Micherdzinski, 1980 Posteroventral spur on coxa III absent...... 8 8. Dorsal shield broad and rounded, peritreme reaching to coxa II, anal shield large and distinctly broad....

Dorsal shield tapering, peritreme extending to coxa I, anal shield smaller and more narrow.....

galagus (Zumpt & Till, 1953)

nyctinomi (Zumpt & Patterson, 1951)

NEW COLLECTION RECORDS

Ornithonyssus bacoti (Hirst, 1913):

Arvicanthis niloticus Parakou, Dahomey 1 female 8 females, 1 PN * Parakou, Dahomey Crocidura sp. 3 males, 12 females Crocidura sp. Borgou region, Dahomey Crocidura flavescens Grahamstown, Cape Province 2 females, 2 PN Crocidura mariquensis Okavango, Botswana 1 male, 3 females Crocidura occidentalis Okavango, Botswana 7 females Dendromus sp. Roodepoort, Transvaal 7 females Herpestes pulverulentis Cape Peninsula, Cape Province 5 females Segbaria, Dahomey Lemniscomys striatus 2 females

^{*} PN = protonymph.

Malacothrix typica Mastomys natalensis Mastomys natalensis Mus musculus Myomys daltoni	Mafeking, Cape Province Soubroukou, Dahomey Panyam Fish Farm, Nigeria Johannesburg, Transvaal Karaduwa, Nigeria	1 2 1	males, 12 females female females female females
Rattus norvegicus	Durban, Natal		females
Rattus rattus	Groot Eiland, Vaaldam, Transvaal		males, 4 females, 8 PN
Rhabdomys pumilio	Namib Desert Park, South West		maios, i Temaios, o 111
- Indiana de la company de	Africa/Namibia	. 8	PN
Ornithonyssus bursa (Berlese, 1888):			
Colius colius	Pretoria, Transvaal	1	female
Laniarus atrococcineus nest	Olifantsfontein, Transvaal	20	females
Myrmecocichla formicivora	Venterstad, Orange Free State	1	female
Native huts	Ovamboland, South West Africa/Na-		
	mibia	10	females
Onychognathus morio nest	Pietermaritzburg, Natal	10	females
Telephorus zeylonus nest	Johannesburg, Transvaal	3	females
Ornithonyssus roseinnesi (Zumpt and	Till, 1953) :		
Aethomys namaquensis	Onseepkans, Cape Province	1	female
Ornithonyssus sylviarum (Canestrini, I	Fanzago, 1877) :		
Myrmecocichla formicivora	East London, Cape Province	1	male, 4 females
Prinia maculosa	Cape Twon, Cape Province	16	females
Pycnonotus barbatus	Pretoria, Transvaal	1	male, 23 females, 3 PN

DISCUSSION

Host and localities for each of the nine species of Ornithonyssus recorded from the Ethiopian region are summarized in Table 2. The majority are endemic to the region, exceptions being O. bacoti, O. bursa and O. sylviarum which are cosmopolitan. These three species are also of medical and veterinary importance. O. bacoti, recorded many times from wild and domestic rodents in Africa, is a common source of human dermatitis. It may also be implicated in the epidemiology of several diseases including plague, tularemia, rickettsialpox and murine typhus (ZUMPT, 1961). The tropical fowl mite, O. bursa, is widely distributed throughout the Ethiopian region and has been recorded from a number of wild bird species and domestic fowl. Similarly, O. sylviarum has been recorded from several wild bird species in the Cape Province and the Transvaal.

Table 2. Hosts and localities of *Ornithonyssus* species in the Ethiopian region.

Species	Locality	Host				
O. africanus (Zumpt & Till, 1958)	Transvaal	Rhinolophys clivosus (Chiroptera)				
O. bacoti (Hirst, 1913)	Cosmopolitan	Domestic and wild rodents				
O. bursa (Berlese, 1888)	Cosmopolitan	Birds				
O. Capensis n. sp.	Cape Province	Elephantulus myurus (In- sectivora)				
O. galagus (Zumpt & Till, 1953)	Uganda/Trans- vaal	Galago senegalensis (Pri- mates)				
		Graphiurus murinus (Ro- dentia)				
 O. lukoschosi Micherd- zinski, 1980 	Zambia	Graphiurus murinus (Ro- dentia)				
O. nyctinomi (Zumpt & Patterson, 1951)	Cape Province	Tadarida bocagei (Chirop- tera)				
O. roseinnesi (Zumpt & Till, 1953)	Cape Province	Rhabdomys pumilio (Ro- dentia)				
		Aethomys namaquensis (Rodentia)				
O. sylviarum (Canestrini & Fanzago, 1877)	Cosmopolitan	Birds				

In general the genus *Ornithonyssus* in the Ethiopian region appears to be adapted to avian

or arboreal hosts, exceptions being O. roseinnesi and O. capensis which have only been recorded from ground-living small mammals. Two species, O. africanus and O. nyctinomi, are parasitic on bats and two, O. bursa and O. sylviarum, exclusively on birds. Both O. lukoschosi and O. galagus have been found on the woodland dormouse (Graphiurus murinus), an arboreal rodent, while O. galagus has also been recorded from the night ape (Galago senegalensis). The primary host of O. bacoti is Rattus rattus which originally may have been arboreal (ROSEVEAR, 1969). Commonly known as the roof rat, it is an excellent climber and it seems probable that O. bacoti was associated with R. rattus prior to its domestication.

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