Acarologia is proudly non-profit,
with no page charges and free open access

Please help us maintain this system by
encouraging your institutes to subscribe to the print version of the journal
and by sending us your high quality research on the Acari.

Subscriptions: Year 2021 (Volume 61): 450 €
http://www1.montpellier.inra.fr/CBGP/acarologia/subscribe.php
Previous volumes (2010-2020): 250 € / year (4 issues)
Acarologia, CBGP, CS 30016, 34988 MONTFERRIER-sur-LEZ Cedex, France
ISSN 0044-586X (print), ISSN 2107-7207 (electronic)

The digitalization of Acarologia papers prior to 2000 was supported by Agropolis Fondation under
the reference ID 1500-024 through the « Investissements d’avenir » programme
(Labex Agro: ANR-10-LABX-0001-01)

Acarologia is under free license and distributed under the terms of the
Creative Commons-BY-NC-ND which permits unrestricted non-commercial use, distribution, and
reproduction in any medium, provided the original author and source are credited.
Contribution to the knowledge of the oribatid mite genus *Perxylobates* (Acari, Oribatida, Haplozetidae)

Sergey G. Ermilov\(^a\), Jhih-Rong Liao\(^b\)

\(^a\) Institute of Environmental and Agricultural Biology (X-BIO), Tyumen State University, Tyumen, Russia.
\(^b\) Department of Entomology, National Taiwan University, Taipei, Taiwan.

Original research

**ABSTRACT**

*Perxylobates hakkai* n. sp. (Oribatida, Haplozetidae) is the second representative of the genus recorded in Taiwan; it is described based on adults from soil under *Ricinus communis* on a farm of the Agricultural Research and Extension Station in Miaoli District. The new species differs from related *Perxylobates coreanus* Choi and Aoki, 1993 by the presence of five pairs of genital setae (versus four pairs), four pairs of notogastral porose areas including A3 (versus three pairs, A3 absent), minute epimeral seta 1c (versus comparatively long), roughened epimeral and aggenital setae (versus heavily barbed), and smooth anterior part of prodorsum (versus foveolate). A revised generic diagnosis and an identification key to known species of *Perxylobates* are provided. The systematic placement of *Perxylobates mahunkai* Bayoumi, 1980 is discussed, resulting in the following new taxonomic proposal: *Protoribates mahunkai* (Bayoumi, 1980) n. comb.

**Keywords** oribatid mites; systematics; morphology; Taiwan

**Zoobank** http://zoobank.org/B053A157-5E77-49B1-AE1D-AA2A18D24C76

**Introduction**

The oribatid mite genus *Perxylobates* of the family Haplozetidae (Acari, Oribatida) was proposed by Hammer (1972) with *Xylobates vermiseta* Balogh and Mahunka, 1968 as type species. According to Subías (online version 2020), the genus comprises 15 species, which are distributed in the Pantropical and Subtropical regions.

During taxonomic identification of oribatid mites from farmland with *Ricinus communis* in Taiwan, we found a new species, belonging to *Perxylobates*. The main goal of our paper is to describe and illustrate this new species. Hitherto, one perylobatid mite species was registered in Taiwan, *P. coreanus* Choi and Aoki, 1993 (Ermilov and Liao 2018).

The additional goals of our paper are to revise generic diagnosis and to provide an identification key to known species of *Perxylobates*, and to discuss the systematic placement of *Perxylobates mahunkai* Bayoumi, 1980.

**Materials and methods**

**Specimens** — Substrate samples containing oribatid mites were collected from the Biological Control Branch, Miaoli District Agricultural Research and Extension Station (Fig. 1) in Taiwan. Mites were extracted from samples into 75% ethanol using Berlese’s funnels with electric lamps in laboratory conditions during 10 days.

As mentioned below in detail, types are distributed among two institutions: the National Taiwan University, Taipei, Taiwan (NTU); the Tyumen State University Museum of Zoology, Tyumen, Russia (TSUMZ).

How to cite this article Ermilov S. G. and Liao J. (2020), Contribution to the knowledge of the oribatid mite genus *Perxylobates* (Acari, Oribatida, Haplozetidae). *Acarologia* 60(3): 612-621; DOI 10.24349/acarologia/20204392
**Observation and documentation** — Specimens were mounted in lactic acid on temporary cavity slides for measurement and illustration. Body length was measured in lateral view, from the tip of the rostrum to the posterior edge of the notogaster. Notogastral width refers to the maximum width of the notogaster in dorsal view (behind pteromorphs). Lengths of body setae were measured in lateral aspect. All body measurements are presented in micrometers. Formulas for leg setation are given in parentheses according to the sequence trochanter-femur-genu-tibia-tarsus (famulus included). Formulas for leg solenidia are given in square brackets according to the sequence genu-tibia-tarsus.

Drawings were made with a camera lucida using a Leica transmission light microscope “Leica DM 2500”.

**Terminology** — Morphological terminology used in this paper follows that of F. Grandjean: see Travé & Vachon (1975) for references, Norton (1977) for leg setal nomenclature, and Norton & Behan–Pelletier (2009), for overview.

**Abbreviations** — Prodorsum: \( \text{lam} \) = lamella; \( \text{slam} \) = sublamella; \( \text{Al} \) = sublamellar porose area; \( \text{tu} \) = tutorium; \( \text{plc} \) = prodorsal lateral carina; \( \text{ro}, \text{le}, \text{in}, \text{bs}, \text{ex} \) = rostral, lamellar, interlamellar, bothридial and exobothridial setae, respectively; \( \text{bos} \) = bothridial scale; \( D \) = dorsophragma; \( P \) = pleurophragma. Notogaster: \( c, \text{la, lm, lp, h, p} \) = notogastral setae; \( Aa, A1, A2, A3 \) = notogastral porose areas; \( ia, im, ip, ih, ips \) = notogastral lyrifissures; \( \text{gla} \) = opisthonotal gland opening. Gnathosoma: \( a, m, h \) = subcapitular setae; \( \text{or} \) = adoral seta; \( d, l, \sup, \inf, \text{cm}, \text{acm, ul, su, lt}, \text{vt} \) = palp setae; \( \omega \) = palp solenidion; \( \text{cha, chb} \) = cheliceral setae; \( \text{Tg} \) = Trägårdh’s organ. Epimeral and lateral podosomal regions: \( 1a, 1b, 1c, 2a, 3a, 3b, 3c, 4a, 4b \) = epimeral setae; \( \text{PdI, PdII} \) = pedotecta I, II, respectively; \( \text{dis} \) = discidium; \( \text{con} \) = concavity; \( \text{cp} \) = circumpedal carina. Anogenital region: \( g, \text{ag, an, ad} \) = genital, aggenital, anal and adanal setae, respectively; \( \text{iad} \) = adanal lyrifissure; \( \text{Amar} \) = marginal porose area; \( \text{po} \) = preanal organ; \( \psi, \tau \) = setae of ovipositor. Legs: \( \text{Tr, Fe, Ge, Ti, Ta} \) = leg trochanter, femur, genu, tibia, tarsus.
respectively; $pa =$ leg porose area; $\omega, \sigma, \varphi =$ leg solenidia; $\varepsilon =$ leg famulus; $d, l, v, ev, bv, ft, tc,$ $it, p, u, a, s, pv, pl =$ leg setae.

**Systematics**

**Generic diagnosis of *Perxylobates***

**Adult** — Sexual species with dimorphism absent. **Body size**: Small to medium (length about 330–550). **Integument**: Body surface smooth or foveolate. **Prodorsum**: Rostrum rounded or slightly truncate. Lamella long, narrow, well separated, without cusp and tooth distally. True translamella absent. Prolamella absent or present. Sublamella and sublamellar porose area present. Tutorium ridge-like. Rostral, lamellar, interlamellar and exobothridial setae developed, usually setiform (rarely ro and/or le thickened), barbed or smooth; ro inserted dorsolaterally on the rostrum, le on end of lamella or on prodorsal surface medial to lam, in interbothridial region. Bothridial seta long, setiform or with unilateral head, ciliate. Bothridium cup-shaped, with anterolateral scale. Dorsosejugal porose area $Ad$ not observed. Dorsosejugal elongate longitudinally. **Notogaster**: Anterior margin of notogaster completely absent. Pteromorph movable, large, curved ventrally. Octotaxic system with three or four (if $A3$ present) pairs of rounded porose areas. With 10 pairs of short, setiform notogastral setae. Dorsosejugal porose area not observed. **Gnathosoma**: Subcapitulum diarthric. Palp with setation 0–2–1–3–9($+$$\omega$). Solenidion of palptarsus connected to eupathidium, located on cylindrical tubercle. Axillary saccule absent. Trägårdh’s organ of chelicera elongate triangular. **Lateral podosomal and epimeral regions**: Pedotecta I and II represented by small lamina. Genal tooth and custodium absent. Discidium and circumpedal carina present. Humeral porose areas $Am$ and $Ah$ not observed. Typical epimeral setal formula 3–1–3–2(or 3). **Anogenital region**: Five (rarely four) pairs of genital, one pair of aggenital, two pairs of anal and three pairs of adanal setae. Adanal seta $ad_1$ posterior, $ad_2$ lateral, $ad_3$ anterior to anal aperture. Adanal lyrifissure located close and lateral to anal plate. Marginal porose area present, narrowly band-like. **Legs**: All legs monodactylous. Porose area present dorsoparaxially on all femora and on trochanters III, IV.

**Juvenile instars** — Not known.

**Perxylobates hakkai** n. sp.

**Diagnosis** — Body size 332–398 × 190–215. Lamella long, about 2/3 length of prodorsum. Rostral, lamellar and interlamellar setae of medium length, setiform, barbed, distance ro–le distinctly shorter than le–in, le pressed to prodorsal surface. Bothridial seta long, with unilaterally dilated head, ciliate. Exobothridial seta minute. Notogastral setae short, setiform, thin, smooth. Four pairs of rounded porose areas present. Epimeral setal formula 3–1–3–2, seta 1e minute, other epimeral and anogenital setae short, setiform, thin, roughened. Five pairs of genital setae. With semi-oval concavity posteriorly to acetabulum IV. Femur III and genu IV with two and one seta, respectively.


**Integument** – Body light brown. Surface of body and all legs microporose (visible in dissected specimens under high magnification, $\times$1000). Antiaxial side of femur III striate.

**Prodorsum** (Figs 2a, 2c) – Rostrum rounded. Lamella long, about 2/3 length of prodorsum. Prolamella absent. Sublamella about 1/3 length of lamella. Sublamellar porose area (18–22 × 10–12) oval, poorly visible. Tutorium of medium size, about 1/3 length of lamella. Prodorsal lateral carina distinct. Rostral (20–28), lamellar (16–24) and interlamellar (24–30) setae setiform, barbed, distance ro–le distinctly shorter than le–in, in erect, le located slightly
Figure 2. *Perxylobates hakkai* n. sp., adult: a – dorsal view (legs omitted); b – ventral view (gnathosoma and legs omitted); c – prodorsum, anterior view; d – lateral view (gnathosoma and legs omitted). Scale bar 50 μm.
Figure 3 *Perxylobates hakkai* n. sp., adult: a– posterior view; b – subcapitulum, ventral view; c – palp, right, antiaxial view; d – chelicera, left, paraxial view; e– ovipositor. Scale bar 50 μm (a), 20 μm (b, d, e), scale bar 10 μm (c).

medial to lamellar end, pressed to prodorsal surface. Bothridial seta (69–77) with long stalk and short, unilaterally dilated head, shortly ciliate. Anterior bothridial scale well developed. Exobothridial seta (2) minute. Sejugal porose area not observed.

Notogaster (Figs 2a, 2c, 3a) – Pteromorph triangular, rounded laterally, with well developed hinge. Ten pairs of notogastral setae (8–10) setiform, thin, smooth. Four pairs of rounded porose areas present, $Aa$ (8–10) larger than $A1–A3$ (4–8), $A3$ located medial to $p_{1}$. All notogastral lyrifissures, opisthognal gland openings and circumgastric scissure distinct. Circumgastric sigillar band not observed.


Epimeral and lateral podosomal regions (Figs 2b, 2c) – Epimeral setae formula 3–1–3–2. Epimeral setae $4c$ absent, $1c$ minute (2), others ($1b$, $3c$ 14–16, $1a$, $2a$, $3a$, $3b$, $4a$, $4b$ 10–12) setiform, thin, roughened. Pedotectum II divided into two scales. Circumpedal carina long, directed to pedotectum II. Discidium triangular. A semi-oval concavity located posteriorly to acetabulum IV.

Anogenital region (Figs 2b, 2c, 3a, 3e) – Five pairs of genital ($g_{1}$ 12, $g_{2}$–$g_{5}$ 8–10), one pair of aggenital (12), two pairs of anal (12) and three pairs of adanal (12) setae setiform, thin, roughened. Adanal lyrifissure distinct. Marginal porose area complete, narrowly band-like. Ovipositor elongated (110 × 41), blade (49) shorter than length of distal section (beyond middle
Figure 4: *Perxylobates hakkai* n. sp., adult: a – leg I, right, antiaxial view; b – leg II, right, antiaxial view; c – leg III, left, antiaxial view; d – leg IV, left, antiaxial view. Scale bar 20 μm.

Each of the three blades with four smooth setae, $\psi_1 \approx \tau_1$ (24–28) setiform, $\psi_2 \approx \tau_2 \approx \tau_3$ (10–12) thorn-like. Six coronal setae not observed.

Legs (Figs 4a-c) – Monodactylous. Claw of all tarsi strong, slightly barbed on dorsal side, with tubercle ventrobasally. Tarsus I with longitudinal ridge distodorsally. Tibiae I and II with tubercle proximodorsally. Dorsoparaxial porose area on femora I-IV and on trochanters III, IV distinct. Proximoventral porose area on all tarsi and distoventral porose area on all tibiae not
Leg setation and solenidia of adult Perxylobates hakkai n. sp.

<table>
<thead>
<tr>
<th>Leg</th>
<th>Tr</th>
<th>Fe</th>
<th>Ge</th>
<th>Ti</th>
<th>Ta</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>v’</td>
<td>d, (l), bv”, v”</td>
<td>(l), v’, σ</td>
<td>(l), (v), ϕ₁, ϕ₂</td>
<td>(l), (tc), (t), (p), (a), (s, pv), v’, (pl), l”, ε, ω₁, ω₂</td>
</tr>
<tr>
<td>II</td>
<td>v’</td>
<td>d, (l), bv”, v”</td>
<td>(l), v’, σ</td>
<td>(l), (v), ϕ</td>
<td>(l), (tc), (t), (p), (a), (s, pv), ω₁, ω₂</td>
</tr>
<tr>
<td>III</td>
<td>v’, l’</td>
<td>d, ev’</td>
<td>l’, σ</td>
<td>l’, (v), ϕ</td>
<td>ft”, (tc), (t), (p), (a), s, (pv)</td>
</tr>
<tr>
<td>IV</td>
<td>v’</td>
<td>d, ev’</td>
<td>d</td>
<td>l’, (v), ϕ</td>
<td>ft”, (tc), (p), (a), (a), s, (pv)</td>
</tr>
</tbody>
</table>

Note: Roman letters refer to normal setae, Greek letters to solenidia (except $ɛ$ = famulus). Single prime (’) marks setae on anterior and double prime (”) setae on posterior side of the given leg segment. Parentheses refer to a pair of setae.

Observe. Formulas of leg setation and solenidia: I (1–5–3–4–20) [1–2–2], II (1–5–3–4–15) [1–1–2], III (2–2–1–3–15) [1–1–0], IV (1–2–1–3–12) [0–1–0]; homology of setae and solenidia indicated in Table 1. Famulus of tarsus I short, erect, slightly dilated apically, inserted between solenidion $ω_2$ and seta $ft’$. Femur III and genu IV with reduced setation, with two and one seta, respectively (seta $l’$ absent). Some ventral setae ($a$, $s$, $pv$) on tarsi I–III thickened, with strong unilateral spines. Solenidion $ω_1$ on tarsus I, $ω_1$ and $ω_2$ on tarsus II and $σ$ on genu III bacilliform, other solenidia setiform.

**Material examined** — Holotype (male) and four paratypes (one male and three females): Taiwan, Miaoli County, Dahu Township, Biological Control Branch, Miaoli District Agricultural Research and Extension Station, 24°25.416' N, 120°52.200' E, 285 m a.s.l., farm, soil under *Ricinus communis* (Fig. 1), 21.III.2018 (J.R. Liao & H.C. Lee).

**Type deposition** — The holotype is deposited in the collection of the NTU; four paratypes are deposited in the collection of the TSUMZ.

**Etymology** — The species name *hakkai* refers to the Hakka. Taiwanese Hakka people is the main population who lives in the type locality Miaoli County, Taiwan.

**Remarks** — Distinctive characters of the new species with the other species of the genus *Perxylobates* can be found in the identification key below.

**Discussion**

*Perxylobates mahunkai* was described by Bayoumi (1980) from Egypt and included in *Perxylobates*. However, this species differs from other representatives of the genus by the presence of a developed anterior notogastral margin (versus anterior notogastral margin completely absent in *Perxylobates* – generic characteristic). All morphological traits of *P. mahunkai* correspond to those of the genus *Protoribates* Berlese, 1908 (e.g. Weigmann et al. 1993; Bayartogtokh 2010), therefore we propose the following combination: *Protoribates mahunkai* (Bayoumi, 1980) **n. comb.**

*Perxylobates mayuloeus* Corpuz-Raros, 1979 from the Philippines was included in *Setoxylobates* Balogh & Mahunka, 1967 by Subías (2004), however, we support its initial position in *Perxylobates* (this was discussed earlier in Ermilov and Liao 2020).

Hence, at present, the genus *Perxylobates* currently comprises 15 species.

**Key to known species of Perxylobates**

We exclude *Perxylobates longissimus* (Warburton, 1912) (Seychelles) from the key because this species has been very briefly and insufficiently described.

1. Bothridial seta with developed head .................................................. 2
   — Bothridial seta setiform .............................................................. 4

— Lamellar seta inserted very close to end of lamella; rostrum rounded .......... 3

3. Five pairs of genital setae; epimal seta lc minute; epimeral and aggenital setae roughened; body size: 332–398 × 190–215 .......... Perxylobates hakkai n. sp. Distribution: Taiwan.

— Epimere I without neotrichy (three pairs of setae) .......... 5

5. Lamellar seta thickened .......... 6
— Lamellar seta setiform .......... 9

— Interlamellar seta of medium length (distinctly shorter than lamellar seta); epimeral setae 1b and 1c not heavily barbed .......... 7

— Insertions of lamellar setae distanced from each other; rostrum rounded .......... 8


— Lamella 1/2 or 2/3 length of prodorsum; tutorium distinctly not reaching rostrum; anterior margin of pteromorph without tooth .......... 10

10. Lamellar seta minute or absent, inserted on end of lamella; all setae of epimere I bushy; body length: 340 .......... Perxylobates siniimes (Hammer, 1971). Distribution: Fiji, Antilles.
— Lamellar seta comparatively long, inserted medial to end of lamella; all setae of epimere I not bushy .......... 11

— All or some setae of epimere I heavily barbed; distance ro-le distinctly shorter than le-in .......... 12

— Interlamellar seta not shorter than lamellar seta. .................................................. 13

— S ubcapitular seta h and all setae of epimere I (1a, 1b, 1c) heavily barbed. .......... 14

14. Lamellar seta distinctly shorter than interlamellar seta; body size: 402 × 205. ...........

References

Acknowledgements

We cordially thank H.C. Lee (National Taiwan University, Taipei, Taiwan) and Miaoli District Agricultural Research and Extension Station for assistance in collecting material; C.C. Ko (National Taiwan University, Taipei, Taiwan) for supporting the study; and Dr. Julia Baumann (University of Graz, Graz, Austria) and two anonymous reviewers for valuable comments.


