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Contribution to the knowledge of the oribatid mite genus *Kalloia* (Acari, Oribatida, Carabodidae), with description of a new species from Indonesia

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Original research

**ABSTRACT**

The genus *Kalloia* (Oribatida, Carabodidae) is recorded in the Oriental region for the first time. A new species — *Kalloia gerdweigmanni* \textit{n. sp.} — is described from litter of oil palm plantations and jungle rubber agroforests from Sumatra, Indonesia. It differs from *Kalloia simpliseta* Mahunka, 1985 by the presence of a transverse ridge in the mediiodistal part of the lamellae, translamella and two thick, diagonal, convergent ridges forming a triangular structure in the medioanterior part of the notogaster, and by the localization of notogastral setae *da*, *dm*, *la*, *lm*, *lp* and *h*\textsubscript{1}. The generic status of *Kalloia* is discussed and supported. *Kalloia mahunkai* Pérez-Íñigo and Baggio, 1989 and *Machadocepheus foveolatus* Mahunka, 1978, which were considered representatives of *Kalloia*, are removed from this genus and combined preliminarily in *Gibbicepheus*. Revised generic diagnosis and data on ecology and distribution of known species of *Kalloia* are presented.

**Keywords** carabodid mites; systematics; morphology; Sumatra; Oriental region

**Zoobank** http://zoobank.org/634891AD-AEA4-4AD4-B188-BA609DDCB287

**Introduction**

The oribatid mite genus *Kalloia* of the family Carabodidae (Acari, Oribatida) was proposed by Mahunka (1985) with *Kalloia simpliseta* Mahunka, 1985 as type species from the Caribbean. He listed the following generic traits: “Prodorsum normal, lamellae simple, running marginally. Lamellar and interlamellar setae thin, simple. Sensillus slightly lanceolate. Dorsosejugal suture present, but a deep hollow lying between prodorsum and notogaster. Latter with some very highly extruding tubercles. Fifteen pairs of thin notogastral setae. Epimeral setal formula: 3–1–3–3. Four pairs of genital, one pair of aggenital, two pairs of anal and three pairs of adanal setae, *ad*\textsubscript{3} in preanal, *ad*\textsubscript{1} and *ad*\textsubscript{2} in postanal position” (Mahunka 1985). According to Subías’s catalogue (2019), the genus is included as subgenus in the genus *Gibbicepheus* Balogh, 1958, and comprises three species (*K. simpliseta, Kalloia mahunkai* Pérez-Íñigo and Baggio, 1989 and *Machadocepheus foveolatus* Mahunka, 1978).

In the course of the study of carabodid mites from Indonesia, we found a new species of *Kalloia*. This paper aims to describe and illustrate this new species under the name *K. gerdweigmanni* \textit{n. sp.}, to update the generic diagnosis, to discuss the taxonomic status of *Kalloia* and the systematic placement of *K. mahunkai* and *M. foveolatus* (to include both species in *Gibbicepheus*), and to present data on the ecology and distribution of *Kalloia* species.
Material and methods

Material

This study was carried out within the framework of the interdisciplinary project “Ecological and socioeconomic functions of tropical lowland rainforest transformation systems (Sumatra, Indonesia)” – EFForTS on Sumatra Island (http://www.uni-goettingen.de/en/310995.html). See the Material examined section for detailed location data of the new species.

In order to study the morphology of the genus Kalloia, besides the new species, the specimens of Kalloia simpliseta from S.G. Ermilov’s collection, from Cuba (Ermilov 2016) and the Caribbean (Ermilov and Smit 2017), were used.

Methods

Specimens were mounted in lactic acid on temporary cavity slides for measurement and illustration of the new species. Body length was measured in lateral view, from the tip of the rostrum to the posterior edge of the notogaster. Notogastral width refers to the maximum width of notogaster in dorsal view. Lengths of body setae were measured in lateral aspect. All body measurements are presented in micrometer. Formulas for leg setation are given according to the sequence trochanter–femur–genu–tibia–tarsus (famulus included). Formulas for leg solenidia are given in square brackets according to the sequence genu–tibia–tarsus. Drawings were made with a camera lucida using a Leica transmission light microscope “Leica DM2500”. SEM photos were made with the aid of a FEI Quanta 250 SEM microscope.

The general morphological terminology used in this paper mostly follows that of F. Grandjean: see Travé and Vachon (1975) for references, Norton (1977) for leg setal nomenclature, and Norton and Behan-Pelletier (2009) for overview.

The following abbreviations are used: lam = lamella; tlam = translamella; r = ridge; dep = depression; tu = tutorium; ro, le, in, bs = rostral, lamellar, interlamellar and bothridial setae, respectively; bo = bothridium; hlp = hump-like process; c, la, lm, lp, h, p = notogastral setae; ia, im, ip, ips = notogastral lyrifissures; gla = opisthonotal gland opening; a, m, h = subcapitular setae; rbr = rutellar brush; v, l, d, cm, acm, ul, sul, vt, lt = palp setae; ω = palp and leg solenidia; cha, chb = cheliceral setae; Tg = Trägårdh’s organ; Pd I, Pd II = pedotecta I and II, respectively; 1a, 1b, 1c, 2a, 3a, 3b, 3c, 4a, 4b, 4c = epimeral setae; dis = discidium; g, ag, an, ad = genital, aggenital, anal and adanal setae, respectively; iad = adanal lyrifissure; p.o. = preanal organ; Tr, Fe, Ge, Ti, Ta = leg trochanter, femur, genu, tibia and tarsus, respectively; p.a. = porose area; σ, φ = legsolenidia; ε = leg famulus; v, ev, bv, l, d, ft, tc, lt, p, u, a, s, pv = leg setae.

Systematics

Generic diagnosis of Kalloia

Dorsoanterior and dorsoposterior parts of notogaster deeply depressed. Dorsocentral part of notogaster with high hump-like process. Fifteen pairs of medium size, setiform notogastral setae widely spaced. Setae  c₁ and c₂ located in anterior notogastral depression; c₃ on humeral shoulders; da, dm, dp, la, lm, lp and h₁ on tubercles/ridges of notogastral hump-like process; five pairs of setae (h₂, h₃, p₁−p₃) in posterior position of notogaster; posterior notogastral depression without setae. Gnathosoma. Subcapitulum diarthric. Adoral setae and theiralveoli absent. Palps with setation 0–2–1–3–9(+ω). Solenidion of palptarsi long, bacilliform, not attached to acm. Axillary saccule absent. Chelicerae chelate-dentate.

**Kalloia gerdweigmanni** n. sp.

_Diagnosis_ — Body size: 531–564 × 298–332. Lamellae mediodistally abruptly bent ventrad, with thicker cuticle appearing as a transverse ridge. Lamellar ends with anterolateral triangular projection. Translamella present. Rostral setae of medium size, thick, spinose. Lamellar setae comparatively short, slightly thickened. Interlamellar setae of medium size, setiform, slightly barbed. Bothridial setae long, setiform, densely barbed. Notogastral setae of medium size, setiform, slightly barbed. Notogastral hump-like process with two medioanterior convergent ridges, forming a triangular structure, bearing setae da and dm; with one pair of medioanterior tubercles, bearing setae dp; with one pair of large, elongate posterolateral tubercles bearing setae la, lm, lp and h₁. Epimeral and anogenital setae (except minute 1a, 1c, la, lm, lp and 2a, 3a and anal setae) setiform, barbed. _Juvenile instars_ — Not known.

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(Figures 1–6)

**Diagnosis** — Body size: 531–564 × 298–332. Lamellae mediodistally abruptly bent ventrad, with thicker cuticle appearing as a transverse ridge. Lamellar ends with anterolateral triangular projection. Translamella present. Rostral setae of medium size, thick, spinose. Lamellar setae comparatively short, slightly thickened. Interlamellar setae of medium size, setiform, slightly barbed. Bothridial setae long, setiform, densely barbed. Notogastral setae of medium size, setiform, slightly barbed. Notogastral hump-like process with two medioanterior convergent ridges, forming a triangular structure, bearing setae da and dm; with one pair of medioanterior tubercles, bearing setae dp; with one pair of large, elongate posterolateral tubercles bearing setae la, lm, lp and h₁. Epimeral and anogenital setae (except minute 1a, 1c, 2a and 3a and anal setae) setiform, barbed.

**Description** — Measurements – Body length: 531 (holotype: female), 531–564 (four paratypes: all females); notogaster width: 307 (holotype), 298–332 (four paratypes).

Integument (Figs 1a, 2a, 3c, 3d, 4a, 4b, 5a, 5c, 5a) – Body color light brown to brown reddish and dark brown. Body covered by thick layer of gel-like cerotegument. Body surface (including subcapitular mentum and genae, genital and anal plates) microtuberculate (diameter of tubercles less than 1). Notogaster and dorsoanaxial part of leg femora III, IV and trochanters III, IV sparsely foveolate (diameter of foveoles up to 12). Projecting parts of lamellae and tutoria slightly foveate.

Prodorsum (Figs 1a, 1b, 2a, 4a, 4b, 5a, 5b, 6a) – Rostrum broadly rounded. Lamellae long (slightly shorter than prodorsum), mediodistally abruptly bent ventrad, with thicker cuticle appearing as a transverse ridge (these regions of lamellae slightly convex, illusory forming the unclear hump-like processes – Figs 4a, 5a, which are really absent – see Figs 4b, 6a), with lateral triangular projection. Translamella broad. Tutoria (3/4 length of prodorsum) strong, ridge-like. With elongate depression between lamellae and tutoria, and two depressions ventrally to tutoria. Rostral setae (41–45) thick, with numerous spines. Lamellar setae (28–32) slightly thickened, roughened, located on translamella. Interlamellar setae (53–61) setiform, thin, slightly barbed. Bothridial setae (77–82) thickened, heavily barbed, curved semi-oval in mediodistal part. Exobothridial setae and their alveoli not observed. Interlamellar region slightly depressed.

Notogaster (Figs 1a, 1b, 2a, 4a, 4b, 5b, 5c, 5a) – Anterior notogastral margin straight. Anterior and posterior notogastral depressions and dorso-central hump-like process well-developed. Medioanterior region of notogastral hump-like process with two thick, diagonal,
**Figure 1** *Kalloia gerdweigmanni* n. sp., adult: a – dorsal view; a – ventral view (gnathosoma and legs omitted). Scale bar 100 μm.

Convergent ridges, forming triangular structure directed in anterior notogastral depression. Medioposterior region of notogastral hump-like process with one pair of tubercles, located behind diagonal ridges; these ridges and tubercles fused or indistinctly connected. Posterolateral regions of notogastral hump-like process with one pair of large, elongate tubercles. Fifteen pairs of notogastral setae (32–36) setiform, thin, slightly barbed; of these, *da* and *dm* located on diagonal notogastral ridges; *dp* on medioposterior tubercles; *la*, *lm*, *lp* and *h₁* on posterolateral tubercles. Lyrifissures and opisthonotal gland openings well visible; *ia* located on humeral shoulders, *im* and *gla* close to each other and anterolateral to posterolateral tubercles, *ip* between *p₁* and *p₂*, *ips* and *ih* on lateral sides of notogaster.


Lateral podosomal and epimeral regions (Figs 1b, 2a, 4b, 5b, 6a) – Pedotecta II rounded in ventral view. Discidia triangular, rounded distally. Two depressions behind acetabula IV. One
pair of large depressions bordered by a strong diagonal ridge. With typical epimeral setation: 3-1-3-3. Epimeral setae 1a, 1c, 2a and 3a minute (4), spiniform, 1b, 3b, 3c, 4a, 4b and 4b (32–36) setiform, barbed; 4b thickest.

Anogenital region (Figs 1b, 2a, 5b, 6b, 5c) – With one pair of long, longitudinal ridges lateral to genital aperture and posterior to epimere IV and several anogenital depressions (one large depression between genital and anal apertures; one pair of small depressions close and posterolateral to genital aperture; one pair of small depressions bearing aggenital setae; one pair of large and indistinct depressions lateral to anal aperture; two pairs of medium depressions posterior to acetabula IV). Usually with three poorly visible short, thin, parallel diagonal furrows lateral to genital aperture. Four pairs of genital, one pair of aggenital and three pairs of adanal setae similar in length (32–36), setiform, slightly barbed. Two pairs of anal setae (8) spiniform. Adanal lyrifissures removed from anal aperture and located lateral to \( ad_3 \). Circumventral ridge poorly developed, interrupted posteriorly.

Legs (Figs 3a-d, 5b, 6a) – Claw of each leg strong, sparsely barbed dorsally and with tooth ventrobasally. Porose area distinct on all femora, not observed on trochanters III, IV. Formulas of leg setation and solenidia: I (1–4–2–4–16) [1–2–2], II (1–4–3–3–15) [1–1–2], III (2–3–1–2–15) [1–1–0], IV (1–2–2–2–11) [0–1–0]; homology of setae and solenidia indicated in Table 1. Famulus of tarsi I short, erect, blunt-ended, inserted posterior to solenidion \( \omega_1 \). Solenidion \( \phi_1 \) on tibiae I very long, setiform; \( \omega_2 \) on tarsi I and \( \phi_2 \) on tibiae I comparatively long, thickened, blunt-ended; other solenidia short (except long \( \omega_2 \) on tarsi I), bacilliform. Dorsoanterior apophysis of tibiae I (bearing \( \phi_1 \)) slightly developed.

**Material examined** — Holotype (female) and two paratypes (two females): Indonesia, Sumatra, Bukit Duabelas landscape, oil palm plantation, research site BO3a, 02°04’15.2”S,
**Figure 3** *Kalloia gerdweigmanni* n. sp., adult: a – leg I, without trochanter, right, antiaxial view; b – femur, genu and tibia of leg II, right, antiaxial view; c – leg III, without tarsus, right, antiaxial view; d – leg IV, left, antiaxial view. Scale bar 50 μm.

102°47’30.6”E, litter, November 2013 (B. Klarner). Two paratypes (two females): Indonesia, Sumatra, Harapan landscape, jungle rubber agroforest, research site HJ4a, 01°47’07.3”S, 103°16’36.9”E, litter, November 2013 (B. Klarner).

**Type deposition** — The holotype is deposited in the collection of LIPI (Indonesian Institute of Science) Cibinong, Indonesia; one paratype is deposited in the collection of the Senckenberg Museum of Natural History, Görlitz, Germany; three paratypes are deposited in the collection of the Tyumen State University Museum of Zoology, Tyumen, Russia. All in ethanol with a drop of glycerol. Additional pictures are available in the online repository www.ecotaxonomy.org.

**Etymology** — The species name is dedicated to our colleague, the well-known acarologist Prof. Dr. Gerd Weigmann (Free University of Berlin, Institute of Zoology, Berlin, Germany), for his extensive contributions to our knowledge of oribatid mites.

**Remarks** — *Kalloia gerdweigmanni* n. sp. differs from the type species of the genus — *Kalloia simpliseta* Mahunka, 1985 — by the presence of a transverse ridge in the mediiodistal part of lamellae (versus ridge on lamellae absent), translamella (versus translamella absent) and two thick, diagonal, convergent ridges, forming a triangular structure in the medioanterior
part of the notogaster (versus notogastral ridges absent), and the localization of each pair of notogastral setae $da$ and $dm$ on one ridge (versus setae located on separate tubercles) and $la$, $lm$, $lp$ and $h_1$ on one large elongate tubercle (versus $la$ and $lm$ on one small tubercle; $lp$ and $h_1$ on another small tubercle).

**Discussion**

Mahunka (1985) proposed the monotypic genus *Kalloia* with *Kalloia simpliseta* from the Caribbean. Later, he confirmed the generic status (e.g., Mahunka 1986, 1998), and this was also supported by other authors (Balogh and Balogh 1988, 1992, 2002; Fujikawa 1991; Ermilov 2016; Ermilov and Smit 2017; Ermilov and N’Dri 2018). Subías (2004) included
Figure 5 *Kalloia gerdweigmanni* n. sp., adult, SEM photos: a – prodorsum dorsal view; b – lateral view; c – notogastral ornamentation and setae *c*₁. Scale bar 50 μm (a), scale bar 200 μm (b), scale bar 20 μm (c).
Figure 6  *Kalloia gerdweigmanni* n. sp., adult, SEM photos: a – laterofrontal view; b – anogenital region, ventral view; c – anoanal region, ventral view. Scale bar 100 μm (a), scale bar 50 μm (b), scale bar 40 μm (c).
**Table 1** Leg setation and solenidia of adult *Kalloia gerdweigmanni* n. sp.

<table>
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<td>v’</td>
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<td>(l), σ</td>
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Note: Roman letters refer to normal setae, Greek letters to solenidia (except ε = famulus). Single prime (’) marks setae on anterior and double prime (””) setae on posterior side of the given leg segment. Parentheses refer to a pair of setae.

*Kalloia* as subgenus in *Diplobodes* Aoki, 1958 and later (2016) in *Gibbicepheus*; Fernandez et al. (2014) included it in *Machadocepeus* (*Kalloia*) Balogh, 1958. The genus *Kalloia* is morphologically similar to *Diplobodes*, *Gibbicepheus* and *Machadocepeus*, however it differs from *Diplobodes* and *Gibbicepheus* by the distinctly depressed anterior part of the notogaster and by the presence of a centrrodorsal notogastral hump-like process (versus anterior part of notogaster not depressed; centrrodorsal part of notogaster without hump-like process); and from *Machadocepeus* by the absence of a centrrodorsal prodorsal hump-like processes on which the interlamellar setae are located.

Also, the genus *Kalloia* is morphologically similar to the genera *Tuberocepeus* Balogh and Mahunka, 1969 and *Mangabebodes* Fernández, Theron, Leiva, Rollard and Tiedt, 2014 in having especially a centrrodorsal notogastral hump-like process and distinctly depressed anterior and posterior parts of the notogaster. However, it differs from both by the presence of 15 pairs of notogastral setae, including the presence of notogastral setae c₁–c₃, and their localization in the depressed anterior part of the notogaster (versus 12 pairs of notogastral setae, setae c₁–c₃ absent, depressed anterior part of the notogaster without setae).

According to recent studies of Carabodidae, the presence or absence of prodorsal and noto-gastral depressions and hump-like processes and localization of interlamellar and notogastral setae are important morphological traits on generic level (Fernandez et al. 2013, 2016, 2018; Ermilov 2018; Ermilov and Starý 2018), therefore we support the generic status of *Kalloia*.

Pérez-Íñigo and Baggio (1989) described the second representative of *Kalloia, Kalloia mahunkai* from Brazil. In addition, Subias (2004) included the species *Machadocepeus foveolatus* from Mauritius in *Diplobodes* (*Kalloia*) and later (2016) in *Gibbicepheus* (*Kalloia*), implying that this species is the third representative of *Kalloia*. However, *K. mahunkai* and *M. foveolatus* do not have anterior notogastral depression and no centrrodorsal notogastral hump-like process (generic traits of *Kalloia*) and correspond to generic traits of the genus *Gibbicepheus*, therefore we suggest to exclude these two species from the genus *Kalloia*, and to combine them in *Gibbicepheus*.

**Distribution and ecology of *Kalloia***

At present, *Kalloia simpliseta* has been recorded from the Neotropical region (Mahunka 1985, 1998; Vázquez-González et al. 2015; Ermilov 2016; Ermilov and Smit 2017) and Côte d’Ivoire (Ermilov and N’Dri 2018); *Kalloia gerdweigmanni* n. sp. is known only from Indonesia (this study). Thus, the genus is distributed in the Neotropical, Ethiopian and Oriental regions.

*Kalloia simpliseta* is widely distributed in the Caribbean. It was described from several localities in Saint Lucia (Mahunka 1985): litter with underlying soil from a natural forest in Micoud, Mahaut, Quillesse Reserve; under bark of coastal trees, accumulated rotten material at tree bases in Anse La Raye, Pilori Pt.; sifted litter, wooden debris from various forest sites in Vigie. Later, the species was recorded by Mahunka (1998) in Saint Lucia from forests in the
vicinity of Halcyon Sands Hotel in Vigie, and by Ermilov and Smit (2017) from litter and soil from different Caribbean islands (Antigua, Grenada, Saint-Barthélémy, Trinidad).

Also, *K. simpliseta* was recorded from tropical ecosystems in Cozumel, Quintana Roo, Mexico (Vázquez-González et al. 2015), from leaf litter of forests in Parque Nacional Alejandro de Humboldt (Ermilov 2016), and from ferrallitic soil of primary forests in Goulrikao, Oumé region, Côte d’Ivoire (Ermilov and N’Dri 2018).

*Kalloia gerdweigmanni* n. sp. is recorded from litter of oil palm plantations and jungle rubber agroforests in Bukit Duabelas and Harapan landscapes, Sumatra, Indonesia.

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Samples collected were based on collection permit no. S.07/KKH-2/2013 issued by the Indonesian Ministry of Forestry. Sample exportation was supported by the Indonesian Institute of Science (register file no. 24/SI/MZB/IV/2014) and based on permit no. 125/KKH-5/TRP/2014 issued by the Ministry of Forestry of the Republic of Indonesia.

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